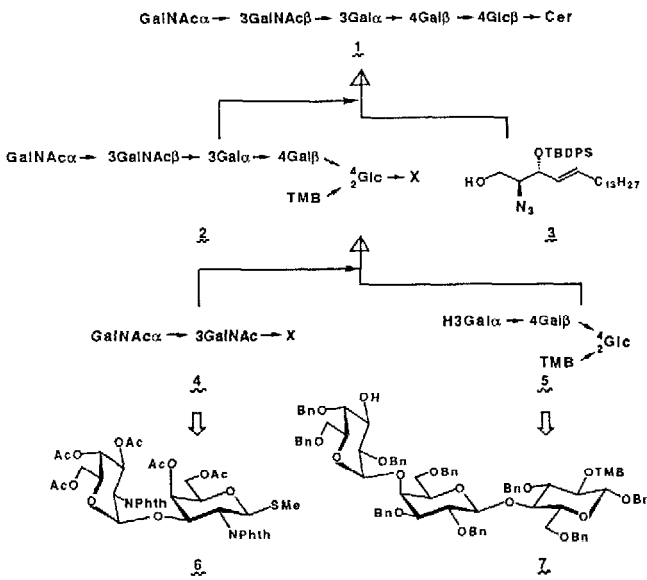


**A TOTAL SYNTHESIS OF FORSSMAN GLYCOLIPID, GLOBOPENTAOSYL CERAMIDE
 IV³GalNAc α Gb₄Cer¹)**

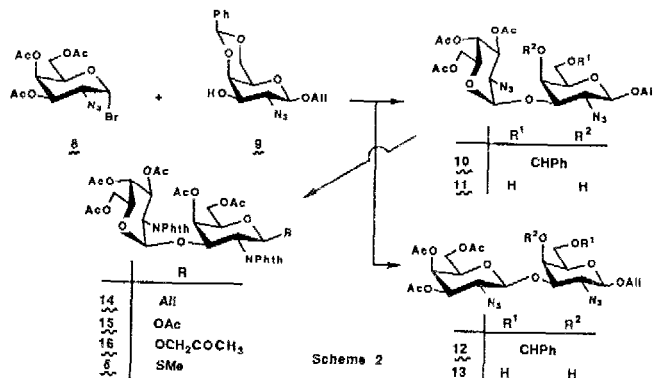
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Abstract: A first total synthesis of Forssman antigen, GalNAc α 1 \rightarrow 3GalNAc β 1 \rightarrow 3Gal α 1 \rightarrow 4Gal β 1 \rightarrow 4Glc β 1 \rightarrow 1Cer, was achieved in a stereoselective manner by using a glycopentaosyl fluoride as a key glycosyl donor for the crucial coupling with a ceramide equivalent.



Scheme 1 (TMB = 2, 4, 6-trimethylbenzoyl, TBDPS = Bu¹Ph₂Si)

A retrosynthetic analysis of **1** led us to design a glycopentaosyl donor **2** that carries a stereocontrolling auxiliary⁸, 2,4,6-trimethylbenzoyl group at O-2a, and a reactive glycosyl acceptor, azido alcohol **3**⁹, as a ceramide equivalent. Compound **2** was further disconnected into a glycobiosyl donor **4** and a glycotriosyl acceptor **5** which were respectively designed as a thioglycoside **6** and a known¹⁰ glycotrioside **7**.

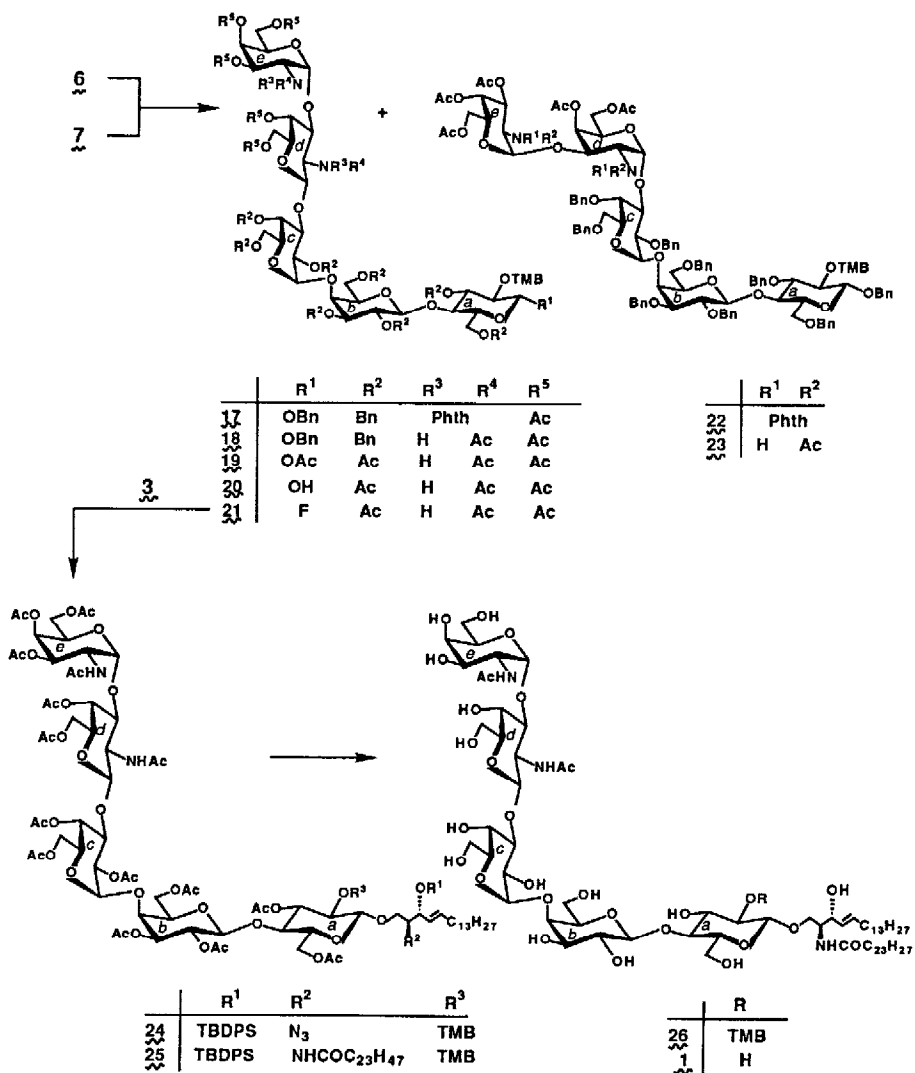


Forssman antigen that occurs² in many species of animals was first described³ in 1911 and was recently reported⁴ as an antigenic marker for a major subpopulation of murine macrophages resided preferentially in spleen and peripheral lymph nodes. The structure of this antigen was proposed⁵ in 1971 as globopentaosyl ceramide **1** from enzymic degradation study. In 1982 first synthesis of glycan part of **1** was achieved⁶ by Paulsen and Bunsch. We now describe a first total synthesis of **1** in a stereocontrolled manner, the ¹H-n.m.r. data of which was found identical with those⁷ of natural sample.

Preparation of **6** was executed in 7 steps from **9**¹¹ in 23% overall yield as follows. AgOTf-Powdered molecular sieves 4A (MS4A) promoted glycosylation of **9** with **8**¹² in (ClCH₂)₂ gave a 94% yield of an

inseparable 5:1 mixture of **10**¹³ and **12**¹³ which was directly solvolized in 4:1 AcOH–H₂O at 80° and separated by a column of silica gel in 1:1 toluene–EtOAc to give **11**¹³ (66%) and **13**¹³ (13%). Conversion of **11** into **14**¹³ was achieved in 3 steps (1 NaOMe in MeOH for 1 h at 20°, 2 HS(CH₂)₃SH¹⁴–Et₃N in MeOH for 12 h at 20°, 3 phthalic anhydride–Et₃N in Py for 2 h at 75°, then Ac₂O was added and 1 h at 75°, 56% overall). Deallylation of **14** with PdCl₂–AcONa in aq. AcOH¹⁵ was found inefficient and gave after acetylation of the products a 1:1 mixture of **15**¹³ and **16**¹³ in 80% yield. The situation was improved by use of a 1:3 mixture of (Ph₃P)₃RhCl–PdCl₂ to give a 70% yield of **15** along with a 29% yield of undesired **16**. Treatment of **15** with Bu₃SnSMe and SnCl₄ in (ClCH₂)₂ for 4 h at 0° afforded thioglycoside **6**¹³ in 95% yield.

Having the designed glycosyl donor **6** prepared, a key glycosylation of the glycotriosyl acceptor **7** with **6** was performed in CH₃NO₂ in the presence of Bu₄NBr–CuBr₂–AgOTf–MS4A¹⁶ for



Scheme 3

13 h at 20° to give a 78% yield of a 10:1 mixture of desired β -D linked glycopentaoside **17**¹³ and the α isomer **22** which were difficult to separate. The mixture was converted into a mixture of acetamido derivatives **18**¹³ and **23**¹³ in 2 steps (1 $\text{NH}_2\text{NH}_2\cdot\text{H}_2\text{O}$ in EtOH for 16 h at 80°, 2 Ac_2O -DMAP in Py, 93% overall) and then separated by a column of silica gel in 2:1 toluene-acetone. Conversion of **18** into a key glycosyl donor **21** which is equivalent to **2** in scheme 1 was smoothly achieved in 4 steps via **19**¹³ and **20**¹³ (1 10% Pd-C and H_2 in 7:3 MeOH- H_2O , 2 Ac_2O -DMAP in Py, 3 $\text{NH}_2\text{NH}_2\cdot\text{AcOH}$ in DMF¹⁷ for 1 h at 20°, 4 DAST¹⁸ in $(\text{ClCH}_2)_2$, 63% overall).

Crucial coupling between the key fluoride **21** and azido alcohol **3** was achieved according to Mukaiyama¹⁹ in the presence of SnCl_2 and AgOTf in $(\text{ClCH}_2)_2$ to give a 23% yield of **24**¹³, which was further converted into the target **1**¹³ in 5 steps via **25**¹³ and **26**¹³ (1 Ph_3P in aq. PhH ²⁰ for 19 h at 45°, 2 $\text{C}_{13}\text{H}_{27}\text{COOH}$ -2-chloro-1-methylpyridinium iodide- Bu_3N ²¹ in $(\text{ClCH}_2)_2$ for 1 h at 20°, 3 Bu_4NF in THF, 4 1:4 0.1M NaOMe-THF for 2 h at 20°, 5 2:1 0.25M NaOMe-THF for 3 h at 60°, 67% overall).

In summary, a stereocontrolled total synthesis of globopentaosyl ceramide **1** was achieved for the first time by employing a thioglycoside **6** and a fluoride **20** as two key glycosyl donors, and the natural sample was unambiguously identified with the synthetic **1** through comparison of their 500 MHz ^1H -n.m.r. data.

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- 13 Physical data for new compounds are compatible with the assigned structures and are given below. Values of $[\alpha]_D$ and $\delta_{\text{H,C}}$ were measured for the solution in CHCl_3 and CDCl_3 , respectively,

at $23 \pm 3^\circ$ unless noted otherwise. **1**: $[\alpha]_D +46.1^\circ$ (c 0.02, Py); δ_H (49:1 $(CD_3)_2SO-D_2O$, 60°) 5.543 (td, 6.7 and 15.3 Hz, 5cer), 5.365 (dd, 6.7 and 15.3 Hz, 4cer), 4.824 (d, 3.7 Hz, 1c), 4.751 (d, 3.6 Hz, 1e), 4.564 (d, 7.8 Hz, 1d), 4.278 (d, 7.8 Hz, 1b), 4.179 (d, 7.8 Hz, 1a), 1.844 and 1.834 (2s, 2NAc), 0.854 (t, 6.7 Hz, 2CH₃). **6**: $[\alpha]_D +137^\circ$ (c 1.0); δ_H 5.961 (dd, 2.8 and 12.2 Hz, 3b), 5.412 (d, 3.7 Hz, 1b), 5.329 (d, 10.3 Hz, 1a); δ_C 94.0 ($^1J_{CH}$ 176 Hz, 1b), 81.1 ($^1J_{CH}$ 156 Hz, 1a). **10 + 12** (5:1 mixture): δ_H 5.593 and 5.581 (2s, in a ratio of 5:1, CHPh), 2.145, 2.050, and 2.044 (3s, 3Ac, **12**), 2.140, 2.063, and 2.038 (3s, 3Ac, **10**). **11**: $[\alpha]_D +86.8^\circ$ (c 1.7); δ_H 5.032 (d, 3.9 Hz, 1b), 4.370 (d, 8.1 Hz, 1a); δ_C 101.5 ($^1J_{CH}$ 160 Hz, 1a), 94.8 ($^1J_{CH}$ 171 Hz, 1b). **13**: $[\alpha]_D +29.0^\circ$ (c 1.4); δ_H 4.574 (d, 7.9 Hz, 1b), 4.357 (d, 7.9 Hz, 1a); δ_C 103.0 ($^1J_{CH}$ 158 Hz, 1b), 101.4 ($^1J_{CH}$ 161 Hz, 1a). **14**: $[\alpha]_D +106^\circ$ (c 0.9); δ_H 5.963 (dd, 3.1 and 12.2 Hz, 3b), 5.394 (d, 3.3 Hz, 1b), 5.324 (d, 8.5 Hz, 1a). **15**: $[\alpha]_D +132^\circ$ (c 1.3); δ_H 6.447 (d, 8.8 Hz, 1a), 5.939 (dd, 2.8 and 12.2 Hz, 3b), 5.412 (d, 3.4 Hz, 1b); δ_C 94.0 ($^1J_{CH}$ 176 Hz, 1b), 90.2 ($^1J_{CH}$ 172 Hz, 1a). **16**: $[\alpha]_D +116^\circ$ (c 1.0); δ_H 6.000 (dd, 3.1 and 12.2 Hz, 3b), 5.388 (d, 3.7 Hz, 1b), 5.271 (d, 8.5 Hz, 1a), 4.183 and 4.116 (2d, 17.4 Hz, OCH₂CO), 2.063, 2.024, 2.007, 2.004, 1.728 and 1.343 (6s, 5Ac and CH₂COMe). **17**: δ_H 6.810 (s, 1.82 H, PhMe₃H₂), 5.900 (dd, 3.0 and 12.1 Hz, 3e), 5.462 (d, 8.0 Hz, 1d), 5.390 (dd, 8.1 and 9.5 Hz, 2a), 5.357 (d, 3.6 Hz, 1e), 4.795 (d, 3.6 Hz, 1c), 4.448 (d, 7.7 Hz, 1b); δ_C 103.0 ($^1J_{CH}$ 163 Hz, 1b), 100.1 ($^1J_{CH}$ 170 Hz, 1c), 99.9 ($^1J_{CH}$ 163 Hz, 1a), 99.7 ($^1J_{CH}$ 163 Hz, 1d), 93.9 ($^1J_{CH}$ 175 Hz, 1e). **22**: δ_H 6.748 (s, 0.18 H, PhMe₃H₂), 5.965 (dd, 3.0 and 12.0 Hz, 3e). **18**: $[\alpha]_D +31.8^\circ$ (c 1.1); δ_H 6.794 (s, PhMe₃H₂), 5.386 (dd, 7.9 and 9.5 Hz, 2a), 5.307 (d, 2.1 Hz, 4e), 5.173 (d, 2.0 Hz, 4d); δ_C 103.0 ($^1J_{CH}$ 163 Hz, 1b), 102.2 ($^1J_{CH}$ 162 Hz, 1d), 100.0 ($^1J_{CH}$ 172 Hz, 1c), 99.9 ($^1J_{CH}$ 162 Hz, 1a), 99.2 ($^1J_{CH}$ 170 Hz, 1e). **23**: $[\alpha]_D +28.9^\circ$ (c 1.0); δ_H 6.820 (s, PhMe₃H₂), 5.366 (dd, 7.8 and 9.5 Hz, H-2a), 5.104 (d, 2.4 Hz, H-4e or d); δ_C 102.92 ($^1J_{CH}$ 163 Hz, 1b), 102.89 ($^1J_{CH}$ 172 Hz, 1d), 101.0 ($^1J_{CH}$ 170 Hz, 1c), 100.0 ($^1J_{CH}$ 163 Hz, 1a), 96.9 ($^1J_{CH}$ 170 Hz, 1e). **19**: δ_H 6.843 (s, PhMe₃H₂), 6.470 (d, 3.7 Hz, 0.5 H, 1a α), 5.711 (d, 8.3 Hz, 0.5 H, 1a β). **20**: δ_H 6.854 and 6.843 (2s, in a ratio of 1:3, PhMe₃H₂). **21**: δ_H 6.860 (s, PhMe₃H₂), 5.880 (dd, 0.3 H, 2.7 and 52.8 Hz, 1a α), 5.472 (dd, 0.7 H, 5.8 and 52.8 Hz, 1a β), 5.611 (d, 0.3 H, 2.8 Hz, 4c α), 5.592 (d, 0.7 H, 3.6 Hz, 4c β). **24**: $[\alpha]_D +46.5^\circ$ (c 0.2); δ_H 6.822 (s, PhMe₃H₂), 6.571 (d, 10.1 Hz, NH), 6.411 (d, 7.0 Hz, NH), 5.606 (d, 3.0 Hz, 4c), 1.050 (s, Bu^t), 0.881 (t, 6.7 Hz, CH₃). **25**: $[\alpha]_D +17.0^\circ$ (c 0.1); δ_H 6.811 (s, PhMe₃H₂), 6.578 (d, 9.7 Hz, NH), 6.550 (d, 8.4 Hz, NH), 5.608 (d, 2.2 Hz, 4c), 5.426 (d, 10.0 Hz, NH), 5.114 (dd, 8.3 and 10.8 Hz, 2a), 1.005 (s, Bu^t), 0.879 (t, 6.6 Hz, 2CH₃). **26**: $[\alpha]_D +32^\circ$ (c 0.05, Py); δ_H (49:1 $(CD_3)_2SO-D_2O$, 60°), 6.854 (s, PhMe₃H₂), 5.518 (td, 7.1 and 15.4 Hz, 5cer), 5.352 (dd, 6.7 and 15.4 Hz, 4cer), 4.849 (t, 9.0 Hz, 2a), 4.793 (d, 3.7 Hz, 1c), 4.731 (d, 4.0 Hz, 1e), 4.552 (d, 8.5 Hz, 1d), 4.544 (d, 8.2 Hz, 1a), 2.232 (s, 3PhMe), 1.826 and 1.831 (2s, 2NAc), 0.843 (t, 7.0 Hz, 2CH₃).

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